

SARS-CoV-2, Flu A, Flu B, RSV Open System PCR Reagents

REF 450-015-Series



24 Reactions 450-015-G-MAX

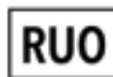
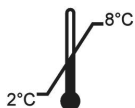


64 Reactions 450-015-H-LMP

Product Insert

For Research Use Only: Not for use in diagnostic procedures

For use with BD MAX™ System, ABI QuantStudio™ 5, ABI 7500 Fast Dx



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For Research Use Only

Research use only reagents are not for use in diagnostic procedures. It is the responsibility of the end user to implement for the intended use.

The SARS-CoV-2, Flu A, Flu B, RSV real-time PCR-based detection reagent is manufactured and packaged as an open system reagent (OSR) for use with open system platforms and has to be validated by the user. Examples of open system platforms are the BD MAX™ System (Windows® Software V4.72A or later), Applied Biosystems QuantStudio™ 5 (Design & Analysis software version 1.5.1 or later) and Applied Biosystems 7500 Fast Dx (SDS software version 1.4 or later).

PLEASE READ ENTIRE PACKAGE INSERT BEFORE PROCEEDING TO USE THE OSR.

PRODUCT OVERVIEW

The BioGX Sample-Ready™ OSR has been formulated in lyophilized format for the multiplex real-time PCR-based detection of RNA from SARS-CoV-2 (N1; nucleocapsid phosphoprotein gene^{1,2,3}), Influenza A (matrix gene⁴), Influenza B (nonstructural gene⁴), RSV A and RSV B (nucleoprotein gene⁴) and a synthetic single stranded RNA Sample Processing Control (SPC) for use with BD MAX extraction kit. The configuration compatible with the ABI platforms detect an integrated synthetic single stranded RNA that serves as an Internal Amplification Control (IAC). The two different formats for the lyophilized Sample-Ready OSR kits are available:

1. BD MAX™ System
REF 450-015-G-MAX
2. ABI QuantStudio™ 5 and Applied Biosystems 7500 Fast Dx Platforms
REF 450-015-H-LMP

Note:

BD MAX™ System OSR (450-015-G-MAX) contains all PCR primers, probes, enzymes, dNTPs, MgCl₂, buffers, and other components required for PCR reaction. BD MAX extraction kits available from Becton, Dickinson and Company include synthetic single stranded RNA sequence which serves as a Sample Processing Control (SPC). No exogenous addition of SPC is needed.

OSR for ABI QuantStudio™ 5 and Applied Biosystems 7500 Fast Dx Platforms (450-015-H-LMP) contain all PCR primers, probes, enzymes, dNTPs, MgCl₂, buffers, and other components required for the PCR reaction. Synthetic single stranded RNA sequence is included in the OSR master mix and serves as Internal Amplification Control (IAC). No exogenous addition of SPC is needed.

PACKAGE CONTENTS

BioGX REF: 450-015-G-MAX

Platform(s): BD MAX™ System

Each 24-reaction package contains two pouches:

1. First pouch contains 24 sealed BD MAX™ tubes of BioGX lyophilized Sample-Ready™ OSR, each tube sufficient for a 12.5 µL PCR reaction.
2. Second pouch contains 24 sealed BD MAX™ tubes, each containing 25 µL of OSR-specific BioGX Rehydration Buffer.

BioGX REF: 450-015-H-LMP

Platform(s): ABI QuantStudio™ 5 and ABI 7500 Fast DX Platforms

Each 64-reaction package contains two pouches:

1. First pouch contains 2 x 8-tube strips. Each tube contains BioGX lyophilized Sample-Ready™ OSR sufficient for 4 x 15 µL PCR reactions.
Note: 8-tube strips containing reagents **ARE NOT** compatible with the ABI QuantStudio™ 5 or ABI 7500 Fast Dx instrument.
2. Second pouch contains 1 vial of BioGX Rehydration Buffer, each containing 750 µL of OSR-specific BioGX Rehydration Buffer (REF 800-0034-HL).

EQUIPMENT AND MATERIALS REQUIRED BUT NOT PROVIDED

- Lyophilized Positive Control Template RNA Beads (10⁵ copies/bead)
 - BioGX SARS-CoV-2 Nucleocapsid phosphoprotein gene (N1) (Part number 720-0206)
 - BioGX Flu A (Part number 720-0002)
 - BioGX Flu B (Part number 720-0003)
 - BioGX RSV A (Part number 720-0181)
 - BioGX RSV B (Part number 720-0182)
- BD MAX™ ExK™ TNA-3 (US BD catalog no. 442827/ International BD catalog no. 442828)
- BD MAX™ PCR Cartridges (US and International BD catalog no. 437519)
- Vortex Genie 2 Vortexer (VWR catalog no. 58815-234) or equivalent
- Disposable nitrile gloves

- Applied Biosystems QuantStudio™ 5 (0.2 mL) consumables
 - Thermo Fisher optical 8-tube strip (catalog no. 4316567)
 - Thermo Fisher ultra-clear optical caps, strips of 8 (catalog no. AB-0866)
 - Thermo Fisher 96-well optical clear reaction plates (catalog no. A36924)
 - Thermo Fisher 96-well qPCR plate seals (catalog no. AB-1170)
- Applied Biosystems 7500 Fast Dx (0.1 mL) consumables.
 - Thermo Fisher optical 8-tube strip (catalog no. 4358293)
 - Thermo Fisher ultra-clear optical caps, strips of 8 (catalog no. 4323032)
 - Thermo Fisher 96-well optical reaction plates (catalog no. 4346906)
 - Thermo Fisher 96-well qPCR plate seals (catalog no. 4311971)

WARNINGS AND PRECAUTIONS

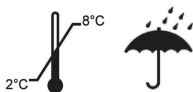


- For research use only. Not intended for use in diagnostic procedures.
- If handling biological samples, including used Extraction Kits and PCR Cartridges, treat as if capable of transmitting infectious agents in accordance with safe laboratory procedures such as those described in CLSI Document M29⁵ and in Biosafety in Microbiological and Biomedical Laboratories⁶.
- BioGX REF: 450-015-G-MAX has been quality control tested only with the BD Open System Extraction Kits on the BD MAX™ System.
- BioGX REF: 450-015-H-LMP has been quality control tested only with the Applied Biosystems QuantStudio™ 5 and Applied Biosystems 7500 Fast Dx real-time PCR platforms.
- Do not use the reagents if the protective pouches are open or torn upon arrival.
- Close reagent protective pouches promptly with the zip seal after each use. Remove any excess air in the pouches prior to sealing and store at 2-8 °C.
- Do not remove desiccant from the PCR master mix pouches.
- Do not use Sample-Ready™ master mix if the desiccant is not present or is broken inside the Sample-Ready™ master mix pouches.
- Do not use reagent tubes if the foil seal has been opened or damaged.
- Do not mix reagents from different pouches and/or kits and/or lots.
- Do not use expired reagents and/or materials.
- Refer to BD MAX™ ExK™ TNA-3 Extraction Kit Instructions or to other respective nucleic acid extraction kits manufacturers' instructions for information about proper handling, cautions, and proper waste disposal.
- Do not mix septum caps between Sample Buffer Tubes or re-use septum caps as contamination may occur and compromise test results.
- Check BD Unitized Reagent Strips for proper liquid fills (ensure that the liquids are at the bottom of the tubes).
- Do not pipette by mouth.
- Do not smoke, drink, or eat in areas where samples or kits are being handled.



- Dispose of unused reagents and waste in accordance with country, federal, provincial, state, and local regulations.
- Use clean gloves when handling extraction kit components and PCR reagents and buffer tubes.

STORAGE REQUIREMENTS AND RECOMMENDATIONS



Reagents are stable at a temperature range of 2-30°C during shipment for 5 days, but BioGX recommends long-term storage at 2-8°C. Reagents have been tested to demonstrate optimal performance when stored properly and consumed by the Manufacturer Recommended Use By Date. The end user may opt to extend the useful life for Research Use Only reagents upon completing their own performance validations. BioGX's guarantee of reagent integrity does not extend beyond the Manufacturer Recommended Use By Date when stored properly. Avoid exposing the reagents (lyophilized or rehydrated) to direct sunlight or long-term ambient lighting. Tightly reseal the pouch with unused reactions and immediately return to a refrigerator after opening. To mitigate reagent performance degradation from exposure to moisture, BioGX suggests using the entire contents of the opened pouch within 1 month; however, the user may choose to verify an extended working time > 1 month by performance testing with positive controls and an examination of the sample preparation control target.

SAMPLE TYPES UTILIZED FOR QUALITY CONTROL TESTING

-Swab samples collected in Copan Universal Transport Media (UTM[®]), BD[™] UVT or saline collections

REAGENT OPTICAL CONFIGURATION

Table 1A. Optical Channel Configuration for REF 450-015-G-MAX.

Optical Channel (Fluorophore Equivalent)	Target
FAM	SARS-CoV-2 (N1)
HEX	Flu A
Texas Red	RSV A/B
Cy5	Flu B
Cy5.5	SPC

Table 1B. Optical Channel Configuration for REF 450-015-H-LMP.

Optical Channel (Fluorophore Equivalent)	Target
FAM	RSV A/B
VIC	SARS-CoV-2 (N1)
TAMRA	Flu B
ROX	Flu A
Cy5	IAC

QUALITY CONTROL AND EXTRACTION TESTING PARAMETERS

BD MAX™ System (BioGX REF: 450-015-G-MAX)

As a starting point, users can import and install the Electronic User Defined Protocol (eUDP) that utilizes **ExK TNA-3** extraction kits onto the BD MAX™. BioGX's most current eUDP utilized for quality control of this product can be obtained by sending an email to TS@biogx.com. Please refer to the BD MAX™ user manual⁷ for uploading instructions.

ABI Platforms (BioGX REF: 450-015-H-LMP)

As a starting point, users can import and install a PCR run file onto:

1. Applied Biosystems QuantStudio™ 5 (Design & Analysis software version 1.5.1 or later)
2. ABI 7500 Fast Dx (SDS software version 1.4 or later)

BioGX's most current ABI QuantStudio™ 5 PCR and ABI 7500 Fast Dx run files utilized for quality control of this product can be obtained by sending an email to TS@biogx.com. Please refer to the Applied Biosystems QuantStudio™ 5 user manual⁸ for uploading instructions. Please refer to the ABI 7500 Fast Dx user manual⁹ for uploading instructions.

SAMPLE VOLUMES UTILIZED DURING QUALITY CONTROL TESTING

The end user may choose to validate a different pretreatment method or volume of sample to load other than the sample processing used by BioGX for QC testing as outlined below.

Swab sample (3 mL Copan Universal Transport Media (UTM®), BD™ UVT or saline collections)

BD MAX™ System

Thoroughly vortex the sample prior to addition to the Sample Buffer Tube (SBT). Pipette 100 µL of sample into the SBT, aseptically place the BD™ septum cap on each SBT. Pulse vortex the SBT for 1-3 seconds, and load the SBT into the extraction tray.

ABI Platforms (validated magnetic bead or silica column extraction kits)

Thoroughly vortex the sample prior to processing. Follow manufacturer recommendations for extraction of appropriate sample volume. Transfer 5 µL to purified nucleic acid to master mix as described in Section: *BioGX Quality Control Test Setup Procedure for ABI Platforms*.

Swab sample (1 mL Copan Universal Transport Media (UTM[®]), BD[™] UVT or saline collections)

BD MAX[™] System

Thoroughly vortex the sample prior to addition to the Sample Buffer Tube (SBT). Pipette 50 µL of sample into the SBT, aseptically place the BD[™] septum cap on each SBT. Pulse vortex the SBT for 1-3 seconds, and load the SBT into the extraction tray.

ABI Platforms (validated magnetic bead or silica column extraction kits)

Thoroughly vortex the sample prior to processing. Follow manufacturer recommendations for extraction of appropriate sample volume. Transfer 5 µL to purified nucleic acid to master mix as described in Section: *BioGX Quality Control Test Setup Procedure for ABI Platforms*.

BioGX QUALITY CONTROL TEST SETUP PROCEDURE

BD MAX[™] System Quality Control Test Setup

Loading a Sample Buffer Tube (SBT)

1. Add the appropriate sample volume to each SBT.
2. Aseptically place BD[™] septum cap on each SBT.
3. Vortex the SBT for 1-3 seconds.
4. Load the SBT into the extraction tray.

WEAR NITRILE GLOVES WHEN HANDLING LYOPHILIZED REAGENTS TO REDUCE THE GENERATION OF STATIC CHARGES. DO NOT USE LATEX GLOVES.

Assembly of BD MAX Extraction Strips and BioGX Reagents

1. Choose the appropriate BD MAX[™] extraction kit (see above). DO NOT use BD MAX[™] master mix or the blank 0.3 mL conical tubes that come with the extraction kit.
2. Load the selected extraction cartridges into the extraction tray, 1 per sample to be tested.
3. Snap one BD MAX[™] ExK[™] TNA Extraction tube into snap-in position 1 (Snap-1) of each extraction strip (**Figure 1**).
4. Snap one BioGX Sample-Ready[™] lyophilized reagent tube into position 2 (Snap-2) of each extraction strip. Check to make sure the lyophilized cake is at the bottom of the tube prior to inserting into the strip. The funnel-shaped cake may be in any orientation (v, >, ^, <).

5. Snap one BioGX Rehydration Buffer tube into position 3 (Snap-3) of each extraction strip. Check to make sure the buffer is at the bottom of the tube prior to inserting into the strip. Position 4 (Snap-4) will remain empty.
6. Lift the tray and briefly examine the bottom of each strip to ensure all reagents are at the bottom of each tube and bubbles are not present in Extraction buffer reservoirs.
7. Proceed with worklist generation and sample loading per BD MAX™ operating instructions. Select the appropriate User Defined Protocol (UDP). Load the extraction tray and, if necessary, a new PCR card into the instrument, close the door, and click “Start Run.”
8. Analyze the results by opening the completed run file in the “Results” tab.

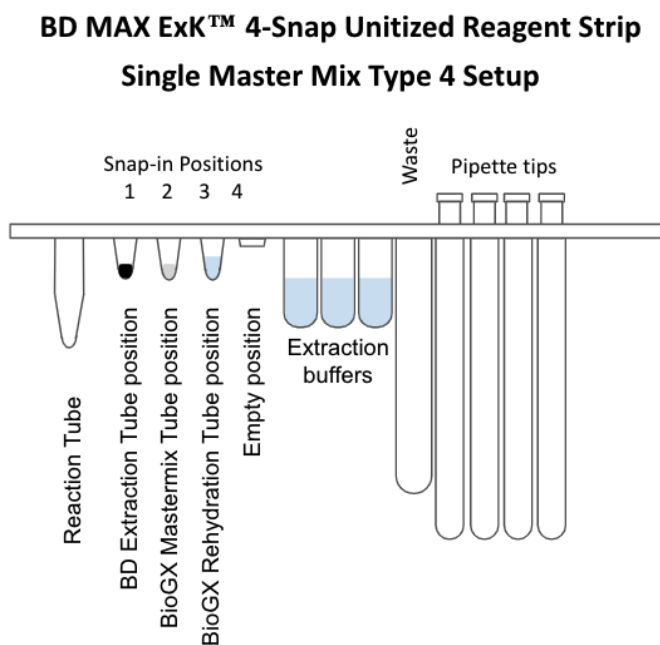


Figure 1. Diagram of BD MAX™ ExK™ 4-snap Unitized Reagent Strips.

Important Note

Always first insert all Snap-1 tubes, then all Snap-2 tubes, then all Snap-3 tubes. The Snap 4 position will remain empty unless the user has set up the reagent to run in dual master mix mode.

Approximately 25 µL of extracted nucleic acid remains in the position 3 tube after extraction. This may be removed and saved for further analyses after the run has been completed.

ABI Platforms Quality Control Test Setup

WEAR NITRILE GLOVES WHEN HANDLING LYOPHILIZED REAGENTS TO REDUCE THE GENERATION OF STATIC CHARGES. DO NOT USE LATEX GLOVES.

Assembly of BioGX Reagents

1. Prepare the appropriate number of 8-tube PCR strips or 96-well PCR plates.
2. Transfer 40 μ L of Rehydration Buffer H to one vial of lyophilized BioGX reagents. The rehydrated master mix is more than sufficient for 4 samples to be tested.

Note: 8-tube strips containing reagents **ARE NOT** compatible with ABI QuantStudio™ 5 or ABI 7500 Fast Dx instrument. Rehydrated master mix must be transferred to 8-tube PCR strips or 96-well PCR plates compatible with the appropriate instrument.

3. Mix by gently pipetting up and down. (IMPORTANT: Keep rehydrated master mix in a cold block or on ice if setup cannot not be completed within 20 minutes. If the rehydrated master mix cannot be used immediately, it can be capped and stored up to 24 hours at 2-8°C, protected from light).
4. Transfer 10 μ L of rehydrated master mix to the bottom of 4 empty wells.
5. To each well containing 10 μ L of rehydrated master mix, add 5 μ L of extracted sample.
6. Affix the appropriate optical caps or optical plate seals.
7. Pulse spin the sealed PCR plate or tube to mix and bring liquid to the bottom.
8. Load 8-tube PCR strips or 96-well PCR plates into the real-time PCR platform and start the run. Avoid unnecessary delay once tubes/plates are loaded into the real-time PCR instrument.

ASSAY PERFORMANCE

All BioGX Research Use Only products are designed to detect 20 copies or less of the target nucleic acid per reaction.

INTERPRETATION OF RESULTS

Table 2A. Multiplex PCR Results Interpretation for 450-015-G-MAX.

SARS-CoV-2	Flu A	RSV A/B	Flu B	SPC	Interpretation
+	-	-	-	+/-	SARS-CoV-2 POSITIVE
-	+	-	-	+/-	Flu A POSITIVE
-	-	+	-	+/-	RSV A/B POSITIVE
-	-	-	+	+/-	Flu B POSITIVE
-	-	-	-	+	SARS-CoV-2, Flu A, RSV A/B, Flu B NEGATIVE
-	-	-	-	-	Unresolved*

*Failed PCR due to inhibition, reagent failure or incorrect assembly of PCR reaction.

Table 2B. Multiplex PCR Results Interpretation for 450-015-H-LMP.

SARS-CoV-2	Flu A	RSV A/B	Flu B	IAC	Interpretation
+	-	-	-	+/-	SARS-CoV-2 POSITIVE
-	+	-	-	+/-	Flu A POSITIVE
-	-	+	-	+/-	RSV A/B POSITIVE
-	-	-	+	+/-	Flu B POSITIVE
-	-	-	-	+	SARS-CoV-2, Flu A, RSV A/B, Flu B NEGATIVE
-	-	-	-	-	Unresolved*

*Failed PCR due to inhibition, reagent failure or incorrect assembly of PCR reaction.









REFERENCES

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<https://www.cdc.gov/coronavirus/2019-ncov/downloads/rt-pcr-panel-primer-probes.pdf>
2. US Centers for Disease Control and Prevention. 2020. Revision to Test Instructions CDC 2019 Novel Coronavirus (nCoV) Real-Time RT-PCR Diagnostic Panel (EUA200001).
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3. US Centers for Disease Control and Prevention. 2020. 2019-Novel coronavirus (2019-nCoV) real-time rRT-PCR panel primers and probes. CDC-006-00019, Revision: 02.
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5. Clinical and Laboratory Standards Institute. Protection of laboratory workers from occupationally acquired infections; Approved Guideline. Document M29 (Refer to the latest edition).
6. Centers for Disease Control and Prevention and National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories. Choosewood L.C. and Wilson D.E. (eds) (2009). HHS Publication No. (CDC) 21-1112.
7. BD MAX™ System User's Manual (refer to the latest revision) BD Life Sciences, Sparks, Maryland 21152 USA.
8. QuantStudio™ Design and Analysis software User Guide, ThermoFisher Scientific, Waltham, Massachusetts, USA (Refer to the latest version).
9. Applied Biosystems 7500 Fast Dx Real-Time PCR Instrument Instructions for Use (2012). Life Technologies Holdings Pte Ltd, Singapore. Publication Part Number 4406991 [Rev. E].

Please call BioGX or email info@biogx.com with any questions you may have regarding this product.

Rev. #	Effective Date	Summary of Changes
06	19 JAN 2026	Update the volume within Buffer HL tube to 750 µL
05	12 NOV 2025	Updated research use only text.
04	03 DEC 2021	Update the product name from "COVID-19, Flu A, Flu B, RSV" to "SARS-CoV-2, Flu A, Flu B, RSV"
03	29 OCT 2021	Update optical channel configuration for 450-015-H-LMP, update rehydration water to rehydration buffer H, remove Bio-Rad CFX96 Touch platform, add ABI 7500 Fast Dx platform
02	04 AUG 2021	Update branding, extraction control for ABI and Bio-Rad platforms and storage conditions.
01	28 MAY 2021	Initial Release.

SYMBOLS

Symbol	Meaning	Symbol	Meaning
	Catalog number		Contains sufficient for <n> tests
	Research Use Only		Manufacturer
	Keep dry		Temperature limitation
	Consult instructions for use		Biological Risks



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